Fermentation extract of *Penicillium italicum* and *Fusarium* oxysporum and a statement of its Biological Effectiveness

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ABSTRACT

The objectives of this study were analysis of the Keywords: Penicillium italicum, Fusarium oxysporum, secondary metabolite products and evaluation of Anti- Fermentation, Biological Effectiveness bacterial activity. The FTIR analysis of Penicillium italicum proved the presence of functional group assignment Alkenes, Alkyl halides, Amide, and Alkane with Intensity (Strong), 1018.41 Stretch (Strong), 819.75 Bending 1238.30 Stretch (Strong), 1379.10 Stretch (Strong), 1614.42 Stretch (Bending), 2850.79 Stretch (Strong), 2920.23Stretch (Strong). The FTIR analysis of Fusarium oxysporum proved the presence of functional group assignment Alkenes, Alkyl halides, Amide, Acid and Alkane with Intensity 875.68 Bending (Strong =C-H), 1016.49 Stretch (Strong C-F), 1024.20 Stretch (Strong C-F), 1197.79 Stretch (Strong C-F), 1317.38 Stretch (Bending N-H), 1716.65 Stretch (Strong C=O), 2850.79 Stretch (Strong C-H), 2922.16 Stretch (Strong C-H) Penicillium italicum bioactive compounds and standard antibiotics were (4.66±0.21), (5.77±0.15), (6.02±0.18), (3.63±0.23), and (5.17±0.32) uses Penicillium italicum bioactive compounds, and (2.11±0.12), (1.25±0.13), (2.98±0.12), (2.00±0.12), and (2.04±0.12) uses Rifambin, and (1.01±0.19), (2.93±0.11), (2.40±0.12), (2.00±0.10), and (1.81±0.11) uses Cefotoxime for Staphylococcus aureus, Escherichia coli, Proteus mirabilis, Klebsiella pneumonia, Pseudomonas eurogenosa respectively. Fusarium oxysporum bioactive compounds standard and antibiotics were (3.07±0.24), (5.94±0.36), (4.18±0.29), (5.63±0.28), and (4.09±0.27) uses *Fusarium oxysporum* bioactive compounds, and (1.09±0.11), (2.89±0.16), (1.59±0.11), (2.25±0.12), and (0.95±0.08) uses Rifambin, and (1.59±0.19), (1.15±0.12), (1.11±0.09), (2.49±0.10), and (0.95±0.07) uses Cefotoxime for Staphylococcus aureus, Escherichia coli, Proteus mirabilis, Klebsiella pneumonia, Pseudomonas eurogenosa respectively

INTRODUCTION

The secondary metabolite may not be directly involved in these activities, but it can have an important and good

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environmental function. For example, resins, terpenes, dyes, and antibiotics. This primary metabolite can be found in many cells or organisms. Also called a central metabolite, it has a more restricted meaning (found in an independently growing organism or any cell) [1-3]. Secondary metabolites also called natural products, specialized metabolites or by-products are organic compounds that are produced by bacteria, fungi or plants that are not directly involved in the development or reproduction of an organism. Secondary metabolites play an important role in plant protection against animal and other defenses between these species. Secondary metabolites are used as drugs and dyes. Secondary host metabolites function in many important functions such as competition and species interactions, except that they are not important for survival. One quality that is necessary for identifying secondary metabolites is their specificity. Fourier transform infrared spectroscopy (FTIR) [4] is a technique that is used to obtain the infrared spectroscopy of the absorption of a solid, gas or liquid substance. The FTIR spectrometer works simultaneously with high resolution data across a large spectral range [5]. This gives a great advantage to a dispersion spectrometer, which operates at intensities over a narrow range of simultaneously. Fourier transform wavelengths spectroscopy is a less intuitive way to obtain the same information. Instead of shining a monochromatic ray of light (a beam of one wavelength) in the sample, this technique illuminates a beam that has many frequencies of light at the same time and measures how much of this beam is absorbed by the sample [6-9]. Then, the beam is modified to contain a different set of frequencies, and this creates a second data point. Also, computer processing is necessary to convert the raw data (light absorption per mirror position) to the desired result (light absorption per wavelength). It turns out that the processing required is a popular algorithm called the Fourier transform. Fourier transform of one field (in this case the displacement of the mirror in centimeters) to its inverse field (wave numbers in 1 cm). This metadata is called a "interference pattern" [10]. Multiplexer or Fellgett Advantage. This starts from that information from all wavelengths is collected. It results in a higher signal-tonoise ratio for a given scanning time limited by the participation of constant detector noise (usually in the infrared thermal spectral region where the photodetector is limited by the generation and recombination noise). The throughput or Jacquinot's advantage. This results from the fact that in a dispersive instrument, the monochromator has entrance and exit slits which restrict the amount of light that passes through it.

MATERIALS AND METHODS

Extraction and purification of the antibacterial agent: Antibacterial compounds were recovered from *Penicillium italicum and Fusarium oxysporum* by solvent extraction with methanol in a ratio of 1:1 (v/v) and shaken well for 1 h. The methanol phase was separated and evaporated to dryness in water bath at $80 - 90^{\circ}$ C. Residue was weighed and re-dissolved with little methanol.

Preparation of sample

About 20 grams of the plant sample powdered were soaked in 100 ml methanol for 16 hours in a rotatory shaker. Whatman No.1 filter paper was used to separate the extract of *Penicillium italicum and Fusarium oxysporum* respectively. The filtrates were used for further phytochemical analysis. It was again filtered through sodium sulphate in order to remove the traces of moisture.

Fourier transform infrared spectrophotometer (FTIR)

The powdered sample of *Penicillium italicum and Fusarium oxysporum* was treated for FTIR spectroscopy (Shimadzu, IR Affinity, Japan). The sample was run at infrared region between 400 nm and 4000 nm [11-13].

Determination of antimicrobial activity of crude bioactive compounds of *Penicillium italicum* and *Fusarium oxysporum*

The test pathogens were swabbed in Müller-Hinton agar plates. Sixty μL of *Penicillium italicum* and *Fusarium oxysporum* extract was loaded on the bored wells. Antibacterial activity was evaluated by measuring the zone of inhibition against the test microorganisms. Methanol was used as solvent control. The antibacterial activity was evaluated by measuring the inhibition-zone diameter observed after 48 h of incubation.

RESULTS AND DISCUSSION

Identification of biochemical compounds

The FTIR analysis of *Penicillium italicum* proved the presence of functional group assignment Alkenes, Alkyl Amide, and Alkane with Intensity 819.75 halides, Bending (Strong), 1018.41 Stretch (Strong), 1238.30 Stretch (Strong), 1379.10 Stretch (Strong), 1614.42 Stretch (Bending), 2850.79 Stretch (Strong). 2920.23Stretch (Strong). The FTIR analysis of Fusarium oxysporum proved the presence of functional group assignment Alkenes, Alkyl halides, Amide, Acid and Alkane with Intensity 875.68 Bending (Strong =C-H), 1016.49 Stretch (Strong C-F), 1024.20 Stretch (Strong C-F), 1197.79 Stretch (Strong C-F), 1317.38 Stretch (Bending N-H), 1716.65 Stretch (Strong C=O), 2850.79 Stretch (Strong C-H), 2922.16 Stretch (Strong C-H). Fusarium oxysporum bioactive compounds and standard antibiotics were (3.07±0.24), (5.94±0.36), (4.18±0.29), (5.63±0.28), and (4.09±0.27) uses Fusarium oxysporum bioactive compounds, and (1.09±0.11), (2.89±0.16), (1.59±0.11), (2.25±0.12), and (0.95±0.08) uses Rifambin, and (1.59±0.19), (1.15±0.12), (1.11±0.09), (2.49±0.10), and (0.95±0.07) uses Cefotoxime for Staphylococcus aureus, Escherichia coli, Proteus mirabilis, Klebsiella pneumonia, Pseudomonas eurogenosa respectively. Infrared spectroscopy provides a useful method for herbal analysis and elucidate the compounds structures as well as for quantitative analysis of drugs. Recently, a number of plants have been reported for antibacterial properties across the world [19-21]. It is hoped that this study would direct to the establishment of some compounds that could be used to invent new and more potent antibacterial drugs of natural origin. Further work will emphasize the isolation and characterization of active principles responsible for bio-efficacy and bioactivity.

able 1. FT-IR peak values of solid analysis of <i>Penicillium italicum</i> .									
No.	Peak (Wave	Intensity	Corr.	Type of	Bond	Type of	Functional		

Group

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	number cm-')		Intensity	Intensity		Vibration	group	frequency	
							assignment		
1.	819.75	80.609	0.316	Strong	=С-Н	Bending	Alkenes	650-1000	
2.	1018.41	61.727	0.876	Strong	C-F	Stretch	alkyl halides	1000-1400	
3.	1238.30	81.739	2.066	Strong	C-F	Stretch	alkyl halides	1000-1400	
4.	1317.38	83.193	0.728	Strong	C-F	Stretch	alkyl halides	1000-1400	
5.	1379.10	81.889	0.178	Strong	C-F	Stretch	alkyl halides	1000-1400	
6.	1614.42	79.693	0.611	Bending	N-H	Stretch	Amide	1550-1640	
7.	2850.79	86.184	4.720	Strong	C-H	Stretch	Alkane	2850-3000	
8.	2920.23	81.949	7.339	Strong	C-H	Stretch	Alkane	2850-3000	
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Table 2. FT-IR peak values of solid analysis of Fusarium oxysporum

No.	Peak (Wave number cm-')	Intensity	Corr. Intensity	Type of Intensity	Bond	Type of Vibration	Functional group assignment	Group frequency
1.	875.68	81.563	1.123	Strong	=С-Н	Bending	Alkenes	650-1000
2.	1016.49	63.828	0.776	Strong	C-F	Stretch	alkyl halides	1000-1400
3.	1024.20	63.550	0.936	Strong	C-F	Stretch	alkyl halides	1000-1400
4.	1197.79	80.489	0.124	Strong	C-F	Stretch	alkyl halides	1000-1400
5.	1317.38	81.051	2.838	Strong	C-F	Stretch	alkyl halides	1000-1400
6.	1614.42	80.833	0.751	Bending	N-H	Stretch	Amide	1550-1640
7.	1716.65	89.259	0.978	Strong	C=0	Stretch	Acid	1700-1725
8.	2850.79	89.239	2.861	Strong	C-H	Stretch	Alkane	2850-3000
9	2922.16	85 898	4 577	Strong	C-H	Stretch	Alkane	2850-3000





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CONCLUSION

The FTIR analysis of *Penicillium italicum* proved the presence of functional group assignment Alkenes, Alkyl halides, Amide, and Alkane. *Penicillium italicum* was very highly active against *Escherichia coli* (6.02 ± 0.18). The FTIR analysis of *Fusarium oxysporum* proved the presence of functional group assignment Alkenes, Alkyl halides, Amide, Acid and Alkane. *Fusarium oxysporum* was very highly active against *Escherichia coli* (5.94 ± 0.36). FTIR analysis of *Penicillium italicum* and *Fusarium oxysporum* revealed the existence of thirty nine bioactive chemical compounds.

REFERENCES

- 1. Hao, W., Li, H., Hu, M., Yang, L. & Rizwan-ul-Haq, M. Integrated control of citrus green and blue mold and sour rot by Bacillus amyloliquefaciens in combination with tea saponin. Postharvest Biol Technol 2011; 59: 316–323
- Kanetis, L., Förster, H. & Adaskaveg, J. E. Determination of natural resistance frequencies in Penicillium digitatum using a new airsampling method and characterization of Fludioxonil- and Pyrimethanil-Resistant isolates. Phytopathology 2010; 100: 738–743
- 3. Azzollini, A. et al. Dynamics of Metabolite Induction in Fungal Co-cultures by Metabolomics at Both Volatile and Non-volatile Levels. Front Microbiol 2018; 9: 72
- 4. Bode, H. B., Bethe, B., Höfs, R. & Zeeck, A. Big efects from small changes: Possible ways to explore nature's chemical diversity. ChemBioChem. 2002; 3: 619–627
- Keller, N. P. Fungal secondary metabolites: regulation, function and drug discovery. Nat Rev Microbiol 2019; 17: 167–180
- Aghcheh, R. K. & Kubicek, C. P. Epigenetics as an emerging tool for improvement of fungal strains used in biotechnology. Appl Microbiol Biotechnol 2015; 99(15): 6167–6181
- 7. Carson, L.A., M.S. Favero, W.W. Bond and N.J. Petersen. Morphological, biochemical and growth characteristics of *Pseudomonas cepacia* from distilled water. Applied Microbiol. 1973; 25: 476-483.
- 8. Charousova, I., J. Medo, E. Halenarova and S. Javorekova. Antimicrobial and enzymatic activity of actinomycetes isolated from soils of coastal islands. J. Adv. Pharm. Technol. Res. 2017; 8: 46-51.

- 9. Cho, K.H., S.T. Kim and Y.K. Kim. Purification of a poreforming peptide toxin, tolaasin, produced by *Pseudomonas tolaasii* 6264. BMB Rep., 40: 113-118.
- Coleman, J.J., S. Ghosh, I. Okoli and E. Mylonakis, 2011. Antifungal activity of microbial secondary metabolites. Plos One. 2007; 6.
- 11. Demain, A.L. and S. Sanchez. Microbial drug discovery: 80 years of progress. J. Antibiot. 2009; 62: 5-16.
- 12. Demain, A.L. Pharmaceutically active secondary metabolites of microorganisms. Applied Microbiol. Biotechnol. 1999; 52: 455-463.
- 13. Desai, D.A., G.P. Kukreja, C.J. Raorane and S.B. Patil. Partial Sequencing of Serendipitously Isolated Antifungal Producer, Pseudomonas tolaasii Strain GD 76 16s Ribosomal RNA Gene. Int. J. Curr. Microbiol. Applied Sci. 2016; 5: 455-458.
- 14. Lu, Y.H., L.P. Jin, L.C. Kong and Y.L. Zhang. Phytotoxic, antifungal and immunosuppressive metabolites from *Aspergillus terreus* QT122 isolated from the gut of dragonfly. Curr. Microbiol. 2017; 74: 84-89.
- Mandryk, M.N., E.I. Kolomiets and E.S. Dey. Characterization of antimicrobial compounds produced by *Pseudomonas aurantiaca* S-1. Polish J. Microbiol. 2007; 56: 245-250.
- 16. Perdicaris, S., T. Vlachogianni and A. Valavanidis. Bioactive natural substances from marine sponges: New developments and prospects for future pharmaceuticals. Nat. Prod. Chem. Res. 2013; 1.
- 17. Pereira, E., A. Santos, F. Reis, R.M. Tavares, P. Baptista, T. Lino-Neto and C. Almeida-Aguiar. A new effective assay to detect antimicrobial activity of filamentous fungi. Microbiol. Res. 2013; 168: 1-5.
- 18. Brimner TA, Boland GJ. A review of the non-target effects of fungi used to biologically control plant diseases. Agr Ecosyst Environ 2003; 100: 3-16.
- 19. El-Tarabily KA, Soliman MH, Nassar AH, Al-Hassani HA, Sivasithamparam K, McKenna F, St J Hardy GE. Biological control of Sclerotinia minor using a chitinolytic bacterium and actinomycetes. Plant Pathol. 2000; 49: 573-83.
- 20. Errakhi R, Bouteau F, Lebrihi A, Barakate M. Evidences of biological control capacities of Streptomyces spp. against Sclerotium rolfsii responsible for damping-off disease in sugar beet (Beta vulgaris L.). World J Microbiol Biotechnol. 2007; 23: 1503-9.

- 21. Xiao K, Kinkel LL, Samac DA. 2002. Biological control of Phytophthora root rots on alfalfa and soybean with Streptomyces. Biol Control. 2002; 23: 285-95.
- 22. Fguira LF, Fotso S, Ameur-Mehdi RB, Mellouli L, Laatsch H. Purification and structure elucidation of antifungal and antibacterial activities of newly isolated Streptomyces sp. strain US80. Res Microbiol. 2005; 156: 341-7.
- 23. Brock, David C."A Measure of Success". Chemical Heritage Magazine. 2011; **29** (1).
- 24. McLafferty, F. W.; Hertel, R. H.; Villwock, R. "Probability based matching of mass spectra. Rapid identification of specific compounds in mixtures". Organic Mass Spectrometry. 1974; 9 (4): 690–702.
- 25. Amirav A, Gordin A, Poliak M, Fialkov AB. "Gas chromatography-mass spectrometry with supersonic molecular beams". J Mass Spectrom. 2008; 43 (2): 141–63.
- 26. Skarysz, A. Convolutional neural networks for automated targeted analysis of raw gas chromatography-mass spectrometry data". International Joint Conferences on Neural Networks. 2018; 1–8.
- 27. Tsivou, M.; Kioukia-Fougia, N.; Lyris, E.; Aggelis, Y.; Fragkaki, A.; Kiousi, X.; Simitsek, P.; Dimopoulou, H.; Leontiou, I. -P.; Stamou, M.; Spyridaki, M. -H.; Georgakopoulos, C. An overview of the doping control analysis during the Olympic Games of 2004 in Athens, Greece". Analytica Chimica Acta. 2006; **555**: 1–13.
- 28. Sloan, K. M.; Mustacich, R. V.; Eckenrode, B. "Development and evaluation of a low thermal mass gas chromatograph for rapid forensic GC-MS analyses". Field Analytical Chemistry & Technology. 2001; 5 (6): 288–301.
- 29. Tekin, Kubilay; Karagöz, Selhan; Bektaş, Sema. A review of hydrothermal biomass processing". Renewable and Sustainable Energy Reviews. 2014; **40**: 673–687.
- 30. Unuh, Mohd Hishamuddin; Muhamad, Pauziah; Waziralilah, Nur Fathiah; Amran, Mohamad Hafiz. "Characterization of Vehicle Smart Fluid using Gas Chromatography-Mass Spectrometry (GCMS)" (PDF). Journal of Advanced Research in Fluid Mechanics and Thermal Sciences. 2019; 55 (2): 240– 248.
- 31. Goesmann, F.; Rosenbauer, H.; Roll, R.; Böhnhardt, H. "COSAC Onboard Rosetta: A Bioastronomy Experiment for the Short-Period Comet 67P/Churyumov-Gerasimenko". Astrobiology. 2005; 5 (5): 622–631.
- 32. Brault, James W. "New Approach to high-precision Fourier transform spectrometer design". Applied Optics. 1996; **35** (16): 2891–2896.
- Smith, D.R.; Morgan, R.L.; Loewenstein, E.V. "Comparison of the Radiance of Far-Infrared Sources". J. Opt. Soc. Am. 1968; 58 (3): 433–434.
- Chamberain, J.; Gibbs, J.E.; Gebbie, H.E. The determination of refractive index spectra by fourier spectrometry". Infrared Physics. 1969; 9 (4): 189–209.
- 35. Brielle, Esther S.; Arkin, Isaiah T. Site-Specific Hydrogen Exchange in a Membrane Environment Analyzed by Infrared Spectroscopy". The Journal of Physical Chemistry Letters. 2018; **9** (14): 4059–4065.